Significant Bacterial Islolates in Patients Presenting with Acute Exacerbation of Idiopathic Pulmonary Fibrosis

Internal Medicine Section

HARISH MAHENDER¹, SURYANARAYANA VEGI², ALEEMULLAH MOHAMMED FAZLUDDIN³, SIVARAJA SUBRAMANIAM⁴

ABSTRACT

Introduction: Idiopathic Pulmonary Fibrosis (IPF) is a chronic and progressive, condition that is due to aberrant wound healing response following repetitive alveolar injury. Though traditionally viruses have played a key role in altering the wound healing cascade, the role played by bacteria in the pathogenesis of IPF is unclear. If there is a recurrent bacterial isolate in sputum or bronchial wash, an appropriate antibiotic both prophylactically and therapeutically can help prevent the decline in lung function and morbidity. This is particularly relevant in the treatment since it involves immunosuppressives during exacerbations.

Aim: This study analyses the sputum for bacterial isolates in patients with acute exacerbation of IPF and aims to find the significance of such association.

Materials and Methods: Sputum samples of 60 patients who came with acute exacerbation of IPF to a Tertiary Medical College and Hospital between February 2015 to March 2016 were analysed. Patients who were unable to produce sputum and who had received antibiotics for the present exacerbation

prior to admission were excluded. Expectorated sputum samples were collected in wide mouth sterile containers and were subjected to gram staining and culture using blood agar after assessing the sputum quality. Chi-Square tests were used to find the significant association between the bacterial isolates in the sputum samples.

Results: Among the 60 patients, there were isolates in 78.3% (n=47) and rest had no isolates. *Streptococcus pneumoniae* was isolated in 12 patients and *Escherichia Coli* in 13 patients and these were found to have significant association statistically in patients presenting with acute exacerbation of IPF. Whereas the rest of the isolates like *Klebsiella* (n=9), *Haemophillus influenza* (n=8), *Enterococcus* (n=2) and *Moraxella catarrhalis* (n=3) were found to have insignificant association.

Conclusion: Preserving the lung functions in patients with IPF is the basis of treatment. Whether infectious causes play a role in attenuating lung function and treating these with prophylactic antibiotics needs to be seen with larger studies in different geographic areas.

Keywords: Blood agar, Lung function, Prophylactic antibiotics, Sputum

INTRODUCTION

The aetiopathogenesis of IPF remains poorly understood. The condition is thought to result from repeated episodes of alveolar injury in individuals with dysfunctional alveolar wound healing mechanism [1]. Though the repetitive alveolar injury and defective healing mechanism plays a vital part in inducing the underlying fibrosis, the environmental factors such as toxins, asbestos, cobalt may cause interstitial pneumonia and fibrosis [2]. Other agents that are known to produce diffuse lung disease include some drugs and radiation. Identified risk factors include exposure to dust, cigarette smoking, gastro oesophageal reflux disease to name a few [3].

Out of the infectious agents, bacterial exacerbation and the possible alteration of alveolar healing by bacteria are the least explored. A recent study by Molyneaux PL et al., has pointed to a disordered host defence and thus susceptibility to infection, as an important contributor to disease progression in IPF [4]. Viruses have long been suspected of playing a role in the pathogenesis of IPF and it is well recognised that many patients report a viral type prodrome preceding the initial development of respiratory symptoms [5].

Acute exacerbations are episodes of rapidly progressive, respiratory decline that occur in individuals with IPF [6]. Histologically, Diffuse Alveolar Damage (DAD) is found in acute exacerbations. Such episodes have been shown in clinical trials to affect between 4-15% of individuals with IPF per year, and are an important cause of IPF related mortality with a 3-month survival of <50% [7,8]. In individuals with IPF, infective exacerbation requiring hospitalisation

confers a mortality risk indistinguishable from that seen with acute exacerbations [9]. In this study we analyse the sputum for bacterial culture in patients who came with acute exacerbation of IPF.

MATERIALS AND METHODS

This was a prospective observational study of sputum samples of 60 patients who came with acute exacerbation of IPF, in Tertiary Medical College, (Sree Balaji Medical College and Hospital, Chennai) between February 2015 to March 2016

Inclusion criteria: All patients who came with acute exacerbation of IPF, diagnosed clinico-radiologically.

Exclusion criteria: Patients who were unable to produce sputum and who had received antibiotic for the present exacerbation prior to admission.

Expectorated sputum samples were collected in wide mouth sterile containers after giving appropriate instructions for all patients. The quality of the sputum was assessed by both macroscopic and microscopic examination. All sputum samples were subjected to Gram staining and culture using blood agar.

STATISTICAL ANALYSIS

Chi-Square tests were used to find the significant association (p-value <0.05) between the bacterial isolates in the sputum samples.

RESULTS

In the study 60 patients were analysed. Among them there were isolates in 78.3% (n=47) and no isolates in 21.7% (n=13) of the subjects.

Among the isolates Streptococcus pneumoniae was isolated in 12 patients and Escherichia coli (E.Coli) in 13 patients. Upon analysis using Chi-square to test the significance of these two isolates, the p value was found to be 0.042 (p<0.05) and 0.032 (p<0.05) for these two isolates respectively, revealing a statistically significant association [Table/Fig-1,2].

	Group			
E.Coli	No isolates (n=13)	Isolates (n=47)	Total	p-value
		n(%)		
No	13 (100.0)	34 (72.3)	47 (78.3)	
Yes	0 (0.0)	13 (27.7)	13 (21.7)	0.032*
Total	13 (100.0)	47 (100.0)	60 (100.0)	
[Table/Fig-1]: Association of <i>E.Coli</i> between isolates and no isolates.				

L			0. 2.00.	
Chi-Square: 2	929	*n<0.05		

	Group				
S. Pneumoniae	No isolates (n=13)	Isolates (n=47)	Total	p-value	
		n(%)			
No	13 (100.0)	35 (74.5)	48 (80.0)		
Yes	0 (0.0)	12 (25.5)	12 (20.0)	0.042*	
Total	13 (100.0)	47 (100.0)	60 (100.0)		
[Table/Fig-2]: Association of <i>S. Pneumoniae</i> between isolates and no isolates. Chi-Souare: 4.149. *p<0.05					

In contrast the other isolates were Klebsiella pneumoniae in 9 patients (p=0.087), Haemophillus influenza (p=0.110), Enterococcus (p=0.449) and Moraxella catarrhalis (0.350) which were found to be statistically insignificant [Table/Fig-3-6].

	Group				
Klebsiella pneumoniae	No isolates (n=13)	Isolates (n=47)	Total	p-value	
		n(%)			
No	13 (100.0)	38 (80.9)	51 (85.0)		
Yes	0 (0.0)	9 (19.1)	9 (15.0)	0.087 (N.S)	
Total	13 (100.0)	47 (100.0)	60 (100.0)		
[Table/Fig-3]: Association of Klebsiella between isolates and no isolates. Chi-Souare: 2.929: N.S: Not significant					

H. Influenza	Group				
	No isolates (n=13)	Isolates (n=47)	Total	p-value	
		n(%)			
No	13 (100.0)	39 (82.9)	52 (86.7)		
Yes	0 (0.0)	8 (17.1)	8 (13.3)	0.110 (N.S)	
Total	13 (100.0)	47 (100.0)	60 (100.0)		
[Table/Fig-4]: Association of <i>H. Influenza</i> between isolates and no isolates.					

	Group			
Enterococcus	No isolates (n=13)	Isolates (n=47)	Total	p-value
	n(%)			
No	13 (100.0)	45 (95.7)	58 (96.7)	
Yes	0 (0.0)	2 (4.3)	2 (3.3)	0.449 (N.S)
Total	13 (100.0)	47 (100.0)	60 (100.0)	

[Table/Fig-5]: Association of Enterococcus between isolates and no isolates. Chi-Square: 0.572; N.S: Not significant

DISCUSSION

In a study done by Molyneaux PL et al., BAL samples were analysed from 25 patients with IPF using culture-independent metagenomic analysis [10]. Their findings were similar to us. In

		Group				
M.Catarrhalis	No isolates (n=13)	Isolates (n=47)	Total	p-value		
		n(%)	<u>.</u>			
No	13 (100.0)	44 (93.6)	57 (95.0)			
Yes	0 (0.0)	3 (6.4)	3 (5.0)	0.350 (N.S)		
Total	13 (100.0)	47 (100.0)	60 (100.0)			
[Table/Fig-6]: Association of <i>M.Catarrhalis</i> between isolates and no isolates. Chi-Square: 0.873; N.S: Not significant						

their study the phylum Firmicutes (Streptococcus and Veillonella species), Proteobacteria, and Bacteroidetes were most commonly encountered. The same authors were able to show by longitudinal analysis of patients with IPF in serum and Bronchoalveolar Lavage (BAL) samples that specific genes, were present in patients with IPF and such expression increased over time, supporting the theory that pathogens may provide chronic antigenic stimuli in patients with IPF [11].

In animal models there is good evidence to show that viral infection can exacerbate already existing fibrosis [12]. There is evidence to suggest a role for viruses in the pathogenesis of IPF, whereas the role of bacteria is less established. One observational evidence comes from Richter AG et al., who in 2008 demonstrated positive BAL cultures in eight of 22 stable IPF patients [13]. A large multicentre, randomized, placebocontrolled study evaluated the prophylactic use of 12 months of cotrimoxazole for IPF [14]. The authors reported that there was no difference in the primary end-point of change in vital capacity when comparing cotrimoxazole and placebo. This observation, together with the high mortality associated with bacterial respiratory tract infection in IPF, suggests that bacteria may play a role in driving IPF disease progression.

However, in a study of 43 individuals suffering an acute exacerbation of IPF, Wootton SC et al., failed to clearly identify a viral or other infectious trigger for the acute exacerbation in the vast majority of their subjects [15]. There were 43 patients with acute exacerbations of IPF and all had negative bacterial cultures and negative viral serology. Subsequent Polymerase Chain Reaction (PCR) analysis of BAL fluid identified four samples positive for rhinovirus, parainfluenza or coronavirus.

Unlike other respiratory conditions where exacerbations are truly acute events, the onset of an acute exacerbation in IPF is generally more insidious [16]. Recently, molecular culture independent techniques have identified complex microbial communities in the lower airways with distinct alterations in the microbiome occurring in a number of respiratory conditions [17,18]. In a study Friaza V et al., where Pneumocystis jirovecii infection was analysed in IPF, a number of uncultured bacteria in the BAL of IPF patients, using basic, culture-independent techniques were identified [19]. He also analysed the microbial flora in the BAL of 20 patients with interstitial lung diseases including idiopathic pulmonary fibrosis, non-specific interstitial pneumonia and acute interstitial pneumonia using gel electrophoresis. Both classic respiratory pathogens (e.g., Haemophilus influenza) and a variety of previously unrecognised or under-recognised organisms were identified. Also, interestingly a negative association was observed between the presence of Pneumocystis jirovecii and bacterial burden, suggesting a possible in vivo antagonism between Pneumocystis and bacterial species.

LIMITATION

The patients who presented with recurrent and similar bacterial isolates need to be followed up and determine whether they have a pattern of colonisation. We did not check the viral and fungal coinfections in these patients.

CONCLUSION

Thus to summarise, in IPF, the role of infectious agents in the pathogenesis and progression of IPF is unknown. Though a variety of bacterial, viral and fungus can be isolated the causal nature of these organisms in IPF remains uncertain. Whether these organisms alter the alveolar healing mechanism and alter the host's response to injury leading to accelerated decline in lung function needs to be seen with long term follow-up studies. Could treating these bacterial infections in IPF patients both with active isolates and with latent infection-prophylactically, reduce mortality and morbidity remains to be seen with a larger study in different settings.

ACKNOWLEDGEMENTS

I thank Tutors India for their statistical help in this study.

REFERENCES

- Maher TM, Wells AU, Laurent GJ. Idiopathic pulmonary fibrosis: multiple causes [1] and multiple mechanisms. Eur Respir J. 2007;30:835-39.
- Du Bois RM. An earlier and more confident diagnosis of idiopathic pulmonary fibrosis. Eur Respir Rev. 2012;21:141-46.
- Taskar VS, Coultas DB. Is idiopathic pulmonary fibrosis an environmental disease. [3] Proc Am Thorac Soc. 2006;3:293-98.
- [4] Molyneaux PL, Willis-Owen SAG, Cox MJ, James P, Cowman S, Loebinger M, et al. Host-microbial interactions in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2017;195:1640-50.
- Golden A, Bronk TT. Diffuse interstitial fibrosis of lungs; a form of diffuse interstitial [5] angiosis and reticulosis of the lungs. AMA Arch Intern Med. 1953;92:106-14.
- [6] Collard HR, Moore BB, Flaherty KR, Brown KK, Kaner RJ, King TE Jr, et al. Acute exacerbations of idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2007;176:636-43.

- [7] Judge EP, Fabre A, Adamali HI, Egan JJ. Acute exacerbations and pulmonary hypertension in advanced idiopathic pulmonary fibrosis. Eur Respir J. 2012:40:93-100.
- [8] Mura M, Porretta MA, Bargagli E, Sergiacomi G, Zompatori M, Sverzellati N, et al. Predicting survival in newly diagnosed idiopathic pulmonary fibrosis: a 3-year prospective study. Eur Respir J. 2012;40:101-09.
- [9] Huie TJ, Olson AL, Cosgrove GP, Janssen WJ, Lara AR, Lynch DA, et al. A detailed evaluation of acute respiratory decline in patients with fibrotic lung disease: aetiology and outcomes. Respirology. 2010;15:909-17.
- Molyneaux PL, Maher TM. Respiratory microbiome in IPF: cause, effect, or [10] biomarker. Lancet Respir Med. 2014;2:511-13.
- [11] Molyneaux PL, Russell AM, Cox MJ, Moffatt MJ, Cookson WOC, Maher TM. The respiratory microbiome in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2012;185:A5174.
- [12] McMillan TR, Moore BB, Weinberg JB, Vannella KM, Fields WB, Christensen PJ, et al. Exacerbation of established pulmonary fibrosis in a murine model by gammaherpesvirus, Am J Respir Crit Care Med. 2008;177;771-80.
- [13] Richter AG, Stockley RA, Harper L, Thickett DR. Pulmonary infection in Wegener granulomatosis and idiopathic pulmonary fibrosis. Thorax. 2009;64:692-97.
- Shulgina L, Cahn AP, Chilvers ER, Parfrey H, Clark AB, Wilson ECF, et al. Treating [14] idiopathic pulmonary fibrosis with the addition of co-trimoxazole: a randomised controlled trial. Thorax, 2013;68:155-62.
- Wootton SC, Kim DS, Kondoh Y, Chen E, Lee JS, Song JW, et al. Viral infection [15] in acute exacerbation of idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2011;183:1698-702
- [16] Cottin V. Changing the idiopathic pulmonary fibrosis treatment approach and improving patient outcomes, Eur Respir Rev. 2012;21:161-67.
- [17] Beasley V, Joshi PV, Singanayagam A, Molyneaux PL, Johnston SL, Mallia P. Lung microbiology and exacerbations in COPD. Int J Chron Obstruct Pulm Dis. 2012;7:555-69
- [18] Hilty M, Burke C, Pedro H, Cardenas P, Bush A, Bossley C, et al. Disordered microbial communities in asthmatic airways. PloS One. 2010;5:e8578.
- [19] Friaza V, la Horra C, Rodríguez-Domínguez MJ, Martín-Juan J, Cantón R, Calderón EJ, et al. Metagenomic analysis of bronchoalveolar lavage samples from patients with idiopathic interstitial pneumonia and its antagonic relation with Pneumocystis jirovecii colonization. J Microbiol Methods. 2010;82:98-101.

PARTICULARS OF CONTRIBUTORS:

- Associate Professor, Department of Respiratory Medicine, Sree Balaji Medical College and Hospital, Bharath University, Chennai, Tamil Nadu, India.
- Professor, Department of Respiratory Medicine, Sree Balaji Medical College and Hospital, Bharath University, Chennai, Tamil Nadu, India. 2. Associate Professor, Department of Microbiology, Madha Medical College and Research Institute, Chennai, Tamil Nadu, India.
- 3. Consultant, Department of Pulmonologist, Delta Care Hospital, Thanjavur, Tamil Nadu, India. 4.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Harish Mahender.

14/18, Third Main Road, Seethammal Colony Alwarpet, Chennai-600018, Tamil Nadu, India. E-mail: drmharish@gmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: Nov 11, 2017 Date of Peer Review: Feb 02, 2018 Date of Acceptance: Apr 21, 2018 Date of Publishing: Jul 01, 2018